

SOME PRACTICAL CONSIDERATIONS IN THE MEASUREMENT OF POLLUTION EFFECTS ON BIVALVE MOLLUSCS, AND SOME POSSIBLE ECOLOGICAL CONSEQUENCES

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A consideration of some physiological (rates of oxygen consumption, the scope for growth) and cellular (the cytochemical latency of a lysosomal enzyme) processes in bivalve molluscs suggests that animal size and seasonal changes related to the gametogenic cycle are important sources of natural variability. Correcting for size using regression techniques, and limiting measurements to one part of the gametogenic cycle, reduces observed natural variability considerably. Differences between populations are then still apparent, but the results of laboratory experiments with hydrocarbons from crude oil suggest that it should be possible to detect sub-lethal effects due to pollution (the 'signal') in the presence of the remaining natural variability (the 'noise'). Statistical considerations, taken together with results from current studies on *Mytilus edulis* and *Scobicularia plana*, indicate that sample sizes of 10-15 individuals should suffice for the detection of possible pollution effects. The physiological effects to be expected in the presence of sub-lethal levels of polluting hydrocarbons are on a scale that can cause significant ecological damage to a population through a reduction in fecundity and the residual reproductive value of the individuals.

Key words: bivalve molluscs, stress responses, natural variability, hydrocarbons, sampling strategy, ecological effects

INTRODUCTION

In measuring the biological effects of pollutants, three questions arise. (1) Can the effect be detected in the environment, amongst the natural variability to be expected between animals in nature? (2) If an effect is evident, what significance has this for the animal's fitness? (3) Can the effect be ascribed to a particular pollutant? Many papers in this Symposium bear on the third of these questions. We wish to consider questions of detection and, more briefly, of ecological significance. We use data obtained by the Stress and Pollution team at I.M.E.R. (see Acknowledgements), working with the common mussel, *Mytilus edulis* L. and the clam *Scrobicularia plana* (da Costa). The various methods employed are described in detail in the original papers referred to in the text.

In considering the detection of a 'pollution effect', two major, natural causes of variance must be considered, viz. the size of the animal and seasonal variability, which is linked to the stage of the gametogenic cycle. Recognizing, and removing, these sources of variance can greatly increase the chance of detecting an effect (physiological or biochemical) due to a pollutant. We discuss some examples and illustrate dose-response relationships from experiments with hydrocarbons. Studies such as these in turn determine the magnitude of difference that must be detected among the natural variance (or 'noise') if a pollution effect is to be established; we illustrate from data the sample sizes that are necessary in such studies. Research into the ecological relevance of such effects is still at an early stage, but some suggestions can be made on the basis of our results.

RESULTS

Two natural causes of variance

Size If individual animals of very similar size (= weight) can be chosen for measurement, the precision in determinations of rates of oxygen consumption (V_{O_2}) and rates of feeding (= clearance rates, CR) can be high. In Table I (line A) V_{O_2} and CR measurements are quoted as means ± 2 SE, together with the coefficient of variation, for a group of 12 *Mytilus* which were selected to be closely matched in shell length and flesh weight. More realistically, a sample with a greater range of individual sizes is normally the best that is available (Table I, line B); less precision is then possible, with coefficients of variation of 20-30% to be expected.

An allometric model of the form:

$$\text{rate} = a \cdot \text{weight}^b,$$

where a and b are fitted parameters, is normally used to describe the relationship between a physiological rate and the weight of the animal. In the statistical treat-

TABLE I

Measurements of the rates of oxygen consumption (V_{O_2} ; ml $O_2 \cdot h^{-1}$) and of clearance rate (CR; litres $\cdot h^{-1}$) in *Mytilus* of mean dry weight 1.66 g. Values are means ± 2 SE, with the coefficient of variation [(SD/mean) $\times 100$] in parentheses. A: animals chosen to be very similar in size; B: animals selected with less attention to similarity in size.

	Dry weight	V_{O_2}	CR
A	1.65 \pm 0.016 (1.6%)	0.431 \pm 0.022 (8.2%)	2.55 \pm 0.34 (21.3%)
B	1.68 \pm 0.193 (18.2%)	0.400 \pm 0.050 (20.0%)	2.56 \pm 0.49 (30.4%)

TABLE II

Analysis of covariance in 14 data sets relating V_{O_2} to body size (dry flesh weight). The F value

Source of variance	d.f.
Total residual variance	238
Differences between slopes	14
Total residual variance	252
Differences between means	14
Total	266

ment of such data the allometric equation is used in a least squares regression analysis; it is difficult to find equations for variation in body size of the form $\text{rate} = \text{weight}^{-b}$.

Such a procedure would be more appropriate if the biological process were constant or nearly constant. It shows the results of regression analysis of V_{O_2} and CR by *Scrobicularia pilosa* (Worrall) between May 1977 and June 1978 (Worrall). In the analysis of covariance, values for b emerged, whereas d values (in the equation) were highly significant (Bayne, 1980). In these circumstances, the data sets and used to reduce the

$$V_{O_2} = W^{-b}.$$

This treatment ascribes no physical meaning to b ; it is, rather, a statistical convenience.

TABLE III

Measurements of the rates of oxygen consumption (V_{O_2} ; ml $O_2 \cdot h^{-1}$) in *Mytilus* of mean dry weight 1.66 g. Values are means ± 2 SE, with the coefficient of variation [(SD/mean) $\times 100$] in parentheses. A: animals chosen to be very similar in size; B: animals selected with less attention to similarity in size.

	Dry weight
A	1.65 \pm 0.016 (1.6%)
B	1.68 \pm 0.193 (18.2%)

TABLE II

An analysis of covariance in 14 data sets relating the rate of oxygen consumption by *Scrobicularia plana* to body size (dry flesh weight). The F value for differences between means is significant at the 1% level.

Source of variance	d.f.	SS	MS	F
Total residual variance	238	6.86	0.029	
Differences between slopes	14	0.45	0.032	1.12
Sub-total residual variance	252	7.31	0.029	
Differences between means	14	10.17	0.719	24.8
Totals	266	17.38		

When such data the allometric equation is usually fitted (on \log_{10} scales) by least-squares regression analysis; it is then possible to 'correct' the physiological determinations for variation in body size by dividing by weight taken to the power b i.e. $\text{rate} \cdot \text{weight}^{-b}$.

Such a procedure would be most useful if values for b for any particular physiological process were constant or at least reasonably similar, over time. Table II shows the results of regression and co-variance analyses of 14 data sets for oxygen consumption by *Scrobicularia plana* measured at seasonally ambient temperatures between May 1977 and June 1979 (data by courtesy of J. Widdows and C.M. Worrall). In the analysis of covariance no significant differences between the fitted values for b emerged, whereas differences between intercepts ($=a$ in the allometric equation) were highly significant (see also Bayne and Widdows, 1978; Newell and Bayne, 1980). In these circumstances a common value for b can be derived for all data sets and used to reduce the variance due to body size as

$$V_{O_2} \cdot W^{-b}$$

This treatment ascribes no particular biological relevance to the fitted value for b ; it is, rather, a statistical convenience to reduce the physiological rate data to a

TABLE III

Measurements of the rates of oxygen consumption (V_{O_2} ; ml $O_2 \cdot h^{-1}$) and of clearance rate (CR; litres $\cdot h^{-1}$) in *Mytilus* of mean dry weight 1.66 g, corrected for variability in body size by use of the fitted parameter b from expressions relating V_{O_2} and CR to dry flesh weight W (see text). A: animals chosen to be very similar in size; B: animals selected with less attention to similarity in size (cf. Table I).

	Dry weight	$V_{O_2} \cdot W^{-0.69}$	CR $\cdot W^{-0.41}$
A	1.65 \pm 0.016 (1.6%)	0.304 \pm 0.016 (8.2%)	2.09 \pm 0.28 (21.5%)
B	1.68 \pm 0.193 (18.2%)	0.279 \pm 0.023 (12.9%)	2.08 \pm 0.30 (22.9%)

major, natural causes of and seasonal variability, recognizing, and removing, the possibility of detecting an effect of hydrocarbons. Studies must be detected to be established; we must research into age, but some suggestions

weight) can be chosen for oxygen consumption (V_{O_2}) in Table I (line A) V_{O_2} and CR with the coefficient of variation to be closely matched in line with a greater range of variation (line B); less precision is to be expected.

to describe the relationship between rate and weight. In the statistical treat-

ment of V_{O_2} and of clearance rate (CR; litres $\cdot h^{-1}$) with the coefficient of variation very similar in size; B: animals

CR
2.55 \pm 0.34
(21.3%)

2.56 \pm 0.49
(30.4%)

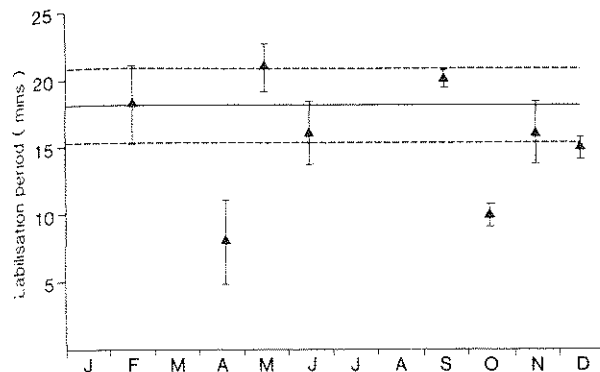


Fig. 2. The labilization period (min) of lysosomal *N*-acetyl- β -hexosaminidase in digestive cells of *Mytilus edulis*; measured under standard conditions over 1 yr. Values are means \pm 95% confidence limits (CL) and the dashed lines indicate the 95% CL of the mean value for the population, excluding the April and October samples.

(Fig. 2) did not differ significantly from a mean of 18 min \pm 0.4 (SE); a total of 40 values in this study had a coefficient of variation of 34%.

Larger seasonal variability is apparent in some physiological processes, such as oxygen consumption rate. These processes tend to be responsive to changes in temperature, salinity, the concentration of suspended particulate matter and oxygen tension. However, here also there is a correlation with the gametogenic stage (Fig. 1). Bayne and Widdows (1978) found that the stage of gametogenic development explained 58% of the variance in measures of V_{O_2} in *Mytilus*, made under field-ambient conditions. Where a full characterization of these seasonal cycles is not feasible, therefore, population comparisons must be made at the same time of year, accepting the increased variance that will result from slight differences in gametogenic condition. Two examples of such a study are briefly considered below.

Variability in the scope for growth and in lysosomal latency

During October, 1978 ten mussels from each of five sites in the Shetland Islands were measured for the suite of physiological processes necessary to the calculation of the scope for growth viz. rate of oxygen consumption, clearance rate, ammonia excretion rate and absorption efficiency. The mussels were chosen to be of similar size and the physiological rates all converted to a weight-specific rate, equivalent to an animal of 1 g dry flesh weight, using exponents fitted by least squares to measurements over a wider weight range made in May, 1978 (Widdows et al., 1981). The results of the scope for growth calculations are shown in Fig. 3. In spite of a wide scatter of points within each population, an analysis of variance indicated significant differences between populations and an *S-N-K* test (Sokal and Rolf, 1969)

edulis related to the state of gametogenesis at an early stage. Coefficient of variation (CV) from 30.4% to

Table I, *b* values of 0.69 (for range of animals. When these to improvement in precision variability (Table III, line A) but a randomly chosen sample

oxygen consumption rate by was examined by a stereology of gametogenic development was 30%; when two the gametogenic cycle (high the gametogenic cycle (low GI), the coefficient. This effect was inde-

the biological response are le at intervals over a year. inidase provides a sensitive 1976, 1980; Bayne et al., 1 yr (Fig. 2) showed two April and in October. These et al., 1981) and experie mussels immediately after, show reduced latency of ll other times of the year

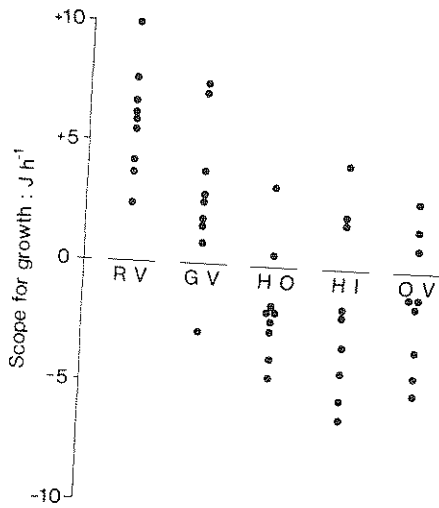


Fig. 3. The scope for growth as $J \cdot h^{-1}$ for *Mytilus edulis* from five populations in the Shetland Islands: RV, Ronas Voe; GV, Gluss Voe; HO, Outer Houb of Scatsta; HI, Inner Houb of Scatsta, OV, Orca Voe. Values are single determinations for $n=9$.

resolved the following ranking:

Ronas Voe > Gluss Voe > Outer Houb = Inner Houb = Orca Voe.

These sites were all relatively unpolluted at the time of the survey. The gametogenic condition of the animals was similar, with the exception of a slightly higher proportion of ripe gametes in individuals from Gluss Voe (Widdows et al., 1981). Judging from these results, therefore, differences between means of $\times 1.5$ to $\times 2.0$ need to be detectable across populations, when measured at the same time of year, if the 'signal' from a potential pollutant is to be detectable.

In a related study on the same five populations of mussels, samples were taken in May 1980 for the assessment of lysosomal latency (Table IV). Measurements were made on five individuals from each site. In the analysis of these data (Table IV) animals from Orca Voe show a slight but insignificant reduction in latency. In order to detect a pollution signal above the natural variability, differences between means of $\times 1.1$ to $\times 1.2$ should suffice.

The effects of hydrocarbons

Bayne et al. (1979), Moore et al. (1980) and Widdows et al. (1981) have reported results from laboratory experiments in which mussels were exposed to low levels of the water-accommodated fraction (WAF) of North Sea crude oil for long periods

TABLE IV

Stabilization period of β -N-acetylhexosaminidase in mussels from the Shetland Islands, U.K.

	Population	
	Ronas Voe	Orca Voe
Stabilization period (min): mean \pm SD for $n=5$	24.0 \pm 5.2	22.0 \pm 4.5
Analysis of variance	d.f.	S
Source of variance		
Between populations	4	1
Within populations	20	4
Total	24	5

and various physiological, cytochemical and histological parameters. In this experiment (not reported in detail) mussels (water without oil) and individuals were measured after 1 and again after 5 weeks. Individuals were carefully selected to be of similar size and scope for growth. In this experiment the ration conditions were constant. There was a depression of the scope for growth

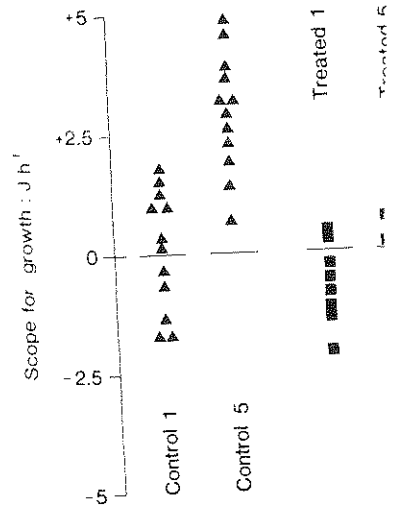


Fig. 4. The scope for growth as $J \cdot h^{-1}$ for mussels exposed to the water-accommodated fraction of North Sea crude oil. Values are single determination

TABLE IV

The labilization period of β -*N*-acetylhexosaminidase in digestive cells of *Mytilus*, measured on five populations from the Shetland Islands, U.K.

	Population				
	Ronas Voe	Gluss Voe	Inner Houb	Outer Houb	Orca Voe
Labilization period (min: mean \pm SD for $n=5$)	24.0 \pm 5.2	25.2 \pm 2.7	22.8 \pm 2.7	22.8 \pm 4.0	19.8 \pm 1.6
Analysis of variance	d.f.	SS	MS	<i>F</i>	
Source of variance					
Between populations	4	80.6	20.2	1.67	n.s.
Within populations	20	241.2	12.1		
Total	24	321.8	—		

and various physiological, cytochemical and biochemical effects observed. In one such experiment (not reported in the papers cited) the scope for growth in control mussels (water without oil) and in mussels exposed to 20–35 $\mu\text{g WAF}\cdot\text{l}^{-1}$ was measured after 1 and again after 5 wk; the sample size was 12, and the mussels were carefully selected to be of similar size at approximately 1 g dry flesh weight. During this experiment the ration conditions improved, resulting in higher values for the scope for growth after 5 wk in both conditions (Fig. 4). Nevertheless, a significant depression of the scope for growth in the WAF-exposed mussels was apparent even

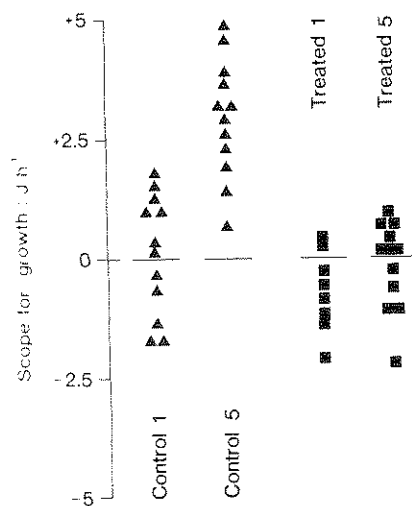


Fig. 4. The scope for growth as $\text{J}\cdot\text{h}^{-1}$ for *Mytilus edulis* exposed to 20–35 $\mu\text{g}\cdot\text{l}^{-1}$ of the water-accommodated fraction of North Sea Crude oil over 1 and 5 wk, compared with controls held in clean sea water. Values are single determinations for $n=12$.

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after 1 wk. As with the results in Fig. 3, the individual scope for growth measurements were variable (low precision) within any one condition, but they showed high sensitivity to the oil. At these low and entirely sub-lethal concentrations of WAF, the 'signal' (i.e. a depression of the scope for growth) was $>100\%$.

The latency of lysosomal hexosaminidase has similarly proved to be sensitive to WAF (Widdows et al., 1981). At a concentration of $7.7 \mu\text{g WAF}\cdot\text{l}^{-1}$, the latency period was reduced from 21.2 ± 3.5 min to 12.5 ± 4.6 min.

These experiments suggest that the effects of low levels of hydrocarbons on *Mytilus* should be detectable in field surveys where the main causes of natural variability i.e. differences in animal size and in gametogenic development, have been controlled or accounted for. The question follows: Is there a reliable dose-response relationship between the biological effect and the pollutant?

The dose-response relationship

Sub-lethal, physiological response studies of the type discussed have not examined the dose-response relationship in detail for any single class of pollutant. The relationship need not be linear. Widdows (1978) determined the scope for growth in *Mytilus* at different combinations of temperature and ration level and described the results by means of multiple regression equations and response surface diagrams; a polynomial expression with 17 terms explained 98% of the variance in 125 data points.

Less is known of the form of relationship to be expected between scope for growth (or lysosomal latency) and the concentration of a pollutant. Some data exist for hydrocarbon effects on bivalves, however; Gillfillan et al. (1976) observed an

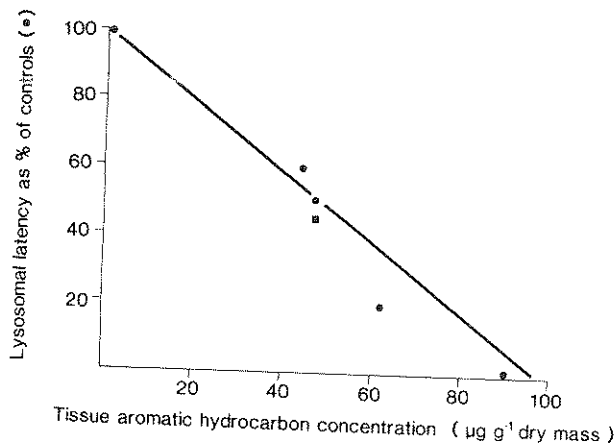


Fig. 5. The relationship between the latency of the lysosomal enzyme *N*-acetyl- β -hexosaminidase (expressed as a % of values in control animals not exposed to hydrocarbons) and the hydrocarbon concentrations in the tissues of *Mytilus edulis*. The data are taken from Widdows et al. (1981).

inverse correlation between carbon aromatic hydrocarbon concentration in *M. edulis*. Widdows et al. (1981) have compared the growth of *Mytilus* and the concentration of hydrocarbons in the tissues of the animals were exposed to WAF. The latency (for hexosaminidase) reduced in the tissue of *Mytilus*, with lat

sample size

Measurements of physiological responses in natural habitats and in laboratory populations that must be detected. On the other hand, knowledge of the seasonal effects, identifies the magnitude of the pollution required size of sample.

Suppose that data are to be compared for a population at two separate time periods. Let y be the rate measurement y (transformed to a normal distribution). (a) Either y is not a function of the pollutant concentration selected. Comparison of the two populations. (b) y is linearly related to the pollutant concentration. This relation is to be estimated. Comparison of the two populations. A t -test would then follow.

(c) The regression lines $y = \beta_0 + \beta_1 x$ are assumed to have common intercepts. Comparison of the two populations. A t -test of the null hypothesis $\beta_0 = \beta_0'$ would then follow.

A further case is as in (b) where the regression lines are assumed to have a common intercept β_0 from previous experience. Comparison of the two populations. A t -test can be defined (see the earlier section).

If any of the above tests are used, the number (n) of observations must be determined by the power of the test. The power is a function of the true difference between the mean y values (of the two populations) and the null hypothesis (of no difference).

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inverse correlation between carbon flux (=scope for growth) and tissue aromatic
hydrocarbon concentration in *Mya arenaria* from sediments polluted by petroleum.
Widdows et al. (1981) have confirmed an inverse correlation between the scope for
growth of *Mytilus* and the concentration of aromatic hydrocarbons in the WAF to
which the animals were exposed in the laboratory. Fig. 5 shows results for lysosomal
latency (for hexosaminidase) related to aromatic hydrocarbon concentration in the
body tissue of *Mytilus*, with latency expressed as a percentage of the control.

Sample size

Measurements of physiological and cytochemical responses by animals in their
natural habitats and in laboratory experiments indicate the differences between
populations that must be detected in any programme of 'effects monitoring'. On the
other hand, knowledge of the natural variance in these responses, due to size and to
seasonal effects, identifies the overall variability within which potential pollutant
effects must be discriminated. These two properties of inherent variance and the
magnitude of the pollution 'signal' are, in turn, the criteria that determine the
required size of sample.

Suppose that data are to be collected from two populations, or from a single
population at two separate times, in order to test for a difference in the mean of a
rate measurement *y* (transformed by taking log₁₀). Three cases are considered:

- (a) Either *y* is not a function of animal size, or animals all of the same size are to be selected. Comparison of the populations is by a simple '2-sample' *t*-test.
- (b) *y* is linearly related to a covariate *x*, for example log₁₀ weight. The slope β of this relation is to be estimated from the data, assuming it is the same for both populations. A *t*-test (equivalently, an *F*-test) of the equality of intercepts would then follow.
- (c) The regression lines $y = \alpha_1 + \beta_1 x$ and $y = \alpha_2 + \beta_2 x$ for the two populations are not assumed to have common slope. All four parameters will be estimated from the data and equality of mean *y* values examined at a specified size *x*₀. This involves a *t*-test of the null hypothesis $(\alpha_2 + \beta_2 x_0) - (\alpha_1 + \beta_1 x_0) = 0$ (see Appendix).

A further case is as in (b) except that β is not estimated from the data; a known value (β_0) from previous experiments is used. 'Corrected' measurements $y' = y - \beta_0 x$ can be defined (see the earlier discussion), and this case is then exactly equivalent to (a).

If any of the above tests is carried out at a fixed significance level $P = 0.05$ or 0.01 , the number (*n*) of observations of *y* that should be taken from each population is determined by the power of the test. This is defined as the probability of rejecting the null hypothesis (of no difference between the population rates) when it is false, that is, when the true difference between the mean *y* values (or intercepts) is $\Delta\alpha$. Power is a function of *n*, *P*, $\Delta\alpha$, σ and *k*, where σ^2 is the usual 'error' variance about the mean *y* values (or regression lines), assumed constant over all observations

from both populations. The factor k reflects expected differences between the animal sizes in the two samples. For the 3 cases above:

- (a) $k = 1$.
- (b) $k^2 = 1 + (\bar{x}_1 - \bar{x}_2)^2 / [2V(x_1) + 2V(x_2)]$, where \bar{x}_1 and $V(x_1)$ denote the mean and variance of the x values in the 1st sample, etc. Thus k will often be close to 1, rising only to $\sqrt{2}$ if the mean of one set of sizes is expected to coincide with an extreme of the other set, and to $\sqrt{5}$ if the two sets are (just) disjoint.
- (c) $k^2 = 1 + [(x_0 - \bar{x}_1)^2 / 2V(x_1)] + [(x_0 - \bar{x}_2)^2 / 2V(x_2)]$. Note that k has roughly the same value here as in (b) if the comparison between the mean y values is made near the point $x_0 = (\bar{x}_1 + \bar{x}_2) / 2$; otherwise k will be larger.

Data from previous laboratory or field experiments are used to supply approximate values for σ (and k). It should be emphasized that such 'guessed' estimates are not used in the analysis ultimately carried out but only in design of the experiment.

Fig. 6 is a set of power curves for $n = 5(1) 10(2) 20(5) 50(10) 100(50) 250$ and $P = 0.05$. (See Appendix for construction details.) Continuous, dashed and dotted lines correspond to cases (a), (b) and (c) respectively. After the first few n values

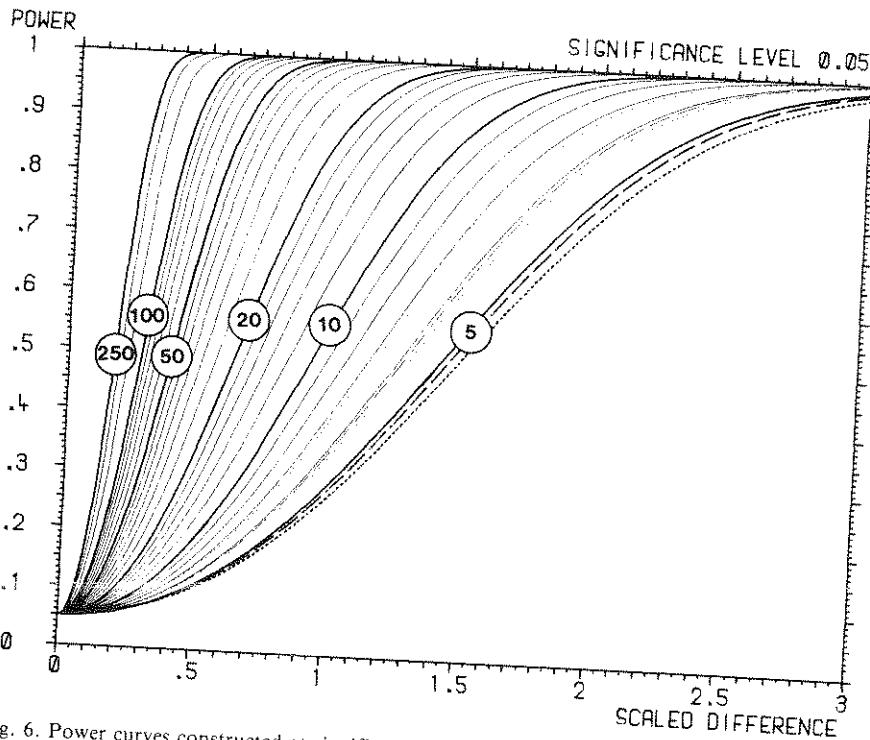


Fig. 6. Power curves constructed at significance level $P = 0.05$ for determining the required sample size for statistical tests described in the text. The circled numbers indicate the sample size ($n = 5 \dots 250$) for three cases discussed in the text: a, solid lines; b, dashed lines; c, dotted lines. At values higher than $n = 7$, the power curves for all three cases are coincident.

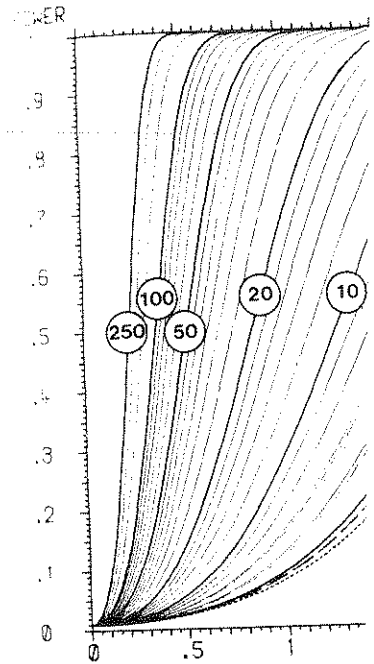


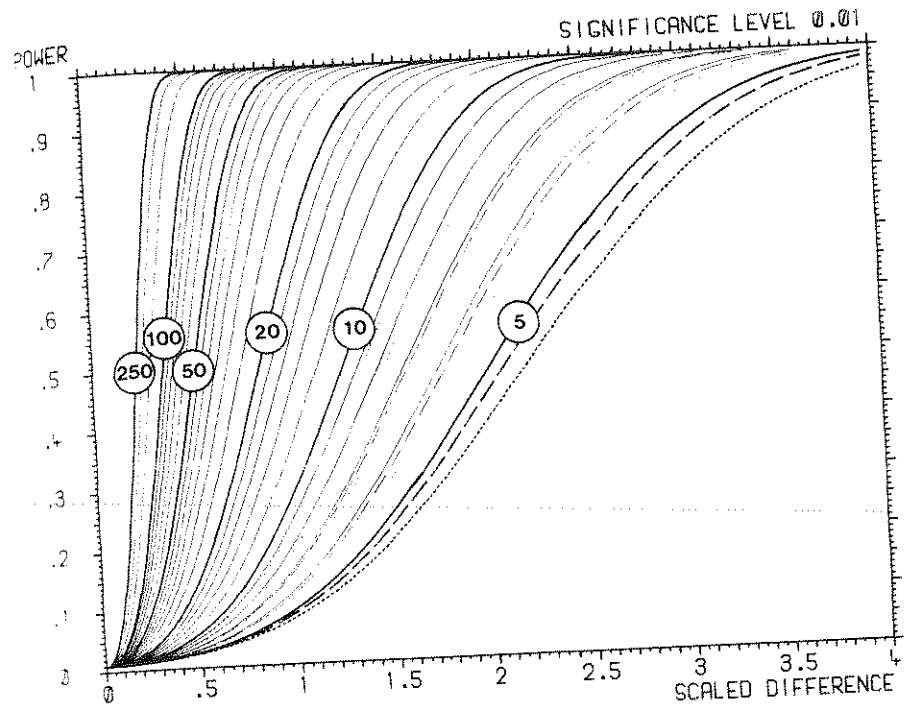
Fig. 7. As for Fig. 6, but with $P = 0.01$.

these lines are coincident and specified mean difference Δ , $(\Delta\alpha)/(k\sigma)$, and for this abscissa proposed tests for various n . It is advisable to use a more stringent test if a large number of samples are to be compared.

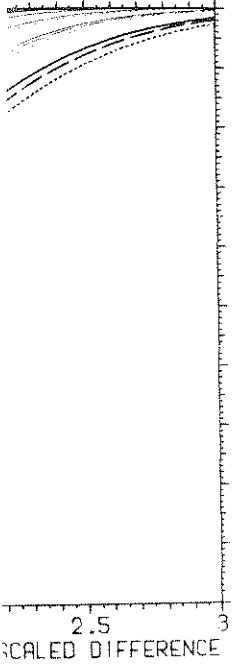
As an example of this procedure, oxygen consumption (e.g. Sc) was measured in a population at two seasons of log₁₀ (oxygen consumption) = 1.5 and 1.6. The difference in mean log₁₀ (oxygen consumption) = 0.1. The residual mean square in the first season (animal weight of 200 mg ($x_0 = 200$)) was 0.1. Also for the second season the difference in mean weights (log₁₀) = 0.150. No assumption is made in the two data sets, so that the required sample size at a power ≥ 0.9

differences between the

denote the mean and will often be close to 1, expected to coincide with an (just) disjoint. that k has roughly the same mean y values is made per. used to supply approximate 'guessed' estimates are design of the experiment.) 50(10) 100(50) 250 and various, dashed and dotted after the first few n values



ANCE LEVEL 0.05



determining the required sample size for the sample size ($n = 5 \dots 250$) lines. At values higher than n

As for Fig. 6, but with $P=0.01$.

These lines are coincident and the continuous lines apply for all 3 cases. For a specified mean difference $\Delta\alpha$ of interest, first calculate the scaled difference $(\Delta\alpha)/(k\sigma)$, and for this abscissa read off ordinates which are the powers of the proposed tests for various n . Fig. 7 provides a similar graph for $P=0.01$. (It is advisable to use a more stringent significance level, like $P=0.01$, if a number of samples are to be compared by t -tests carried out on all possible pairings.)

As an example of this procedure, we consider a programme to measure rates of oxygen consumption (e.g. *Scrobicularia plana*, see Table II) by individuals from one population at two seasons of the year (winter and summer). In the regressions of \log_{10} (oxygen consumption) = $\alpha_1 + \beta_1 \log_{10}$ (dry weight), we wish to detect a difference in mean \log_{10} (oxygen consumption) values of 0.2, at $P=0.01$, for a mean animal weight of 200 mg ($x_0 = 2.30$). From previous experience we accept a value for the residual mean square in the regression analysis ($\log \dot{V}_{O_2}$ against $\log W$) of 0.01; therefore equals 0.1. Also from previous experience we are able to predict a difference in mean weights (\log_{10}) of 0.200 with a variance in the weight measurements of 0.50. No assumption is made as to equality of slopes (β) in the regression analysis of the two data sets, so that the case (c) (above) is used to determine the required sample size at a power ≥ 0.9 .

Using the equations given: $\Delta\alpha = 0.200$; $\sigma = 0.100$; $k = 1.03$. The scaled difference $[(\Delta\alpha)/(k\sigma)] = 1.94$ and (from Fig. 7) a sample size of $n = 10$ would provide a test with power = 0.9.

ECOLOGICAL CONSEQUENCE

We posed the question as to the ecological significance of an observed response to a pollutant, implying that unless the response has a damaging ecological consequence pollution cannot truly be said to occur. With the kinds of data discussed in this paper in mind, the question is more succinctly put: What is the ecological significance of a reduction in the scope for growth?

A decline in the scope for growth, under conditions of constant ration, signifies an impaired growth efficiency, and an inevitable result will be the smaller size of individuals. *Mytilus*, in common with most bivalves, increases egg production with increase in size (and concomitant advance in age). Thompson (1979) fitted allometric equations to data relating weight loss on spawning (an index of fecundity) to dry flesh weight of *Mytilus* from three populations in North America; in all cases the weight exponent was > 1.0 . Bayne and Worrall (1980) recorded weight exponents of 1.40 and 1.29 for spawning weight losses in two populations in the U.K. These values > 1.0 signify an acceleration in the allocation of resources to gametes as the mussels grow in size. Reduced growth efficiency, therefore, lowers the fitness of the individual by indirectly reducing fecundity.

Evidence from laboratory experiments suggests that environmental stress may also affect fecundity directly. In experiments reported by Bayne et al. (1975, 1978), mussels were held at temperatures and rations designed to force a reduction in the scope for growth. The mussels were then induced to spawn and the eggs released were counted (Fig. 8A); the eggs were also analysed for protein, lipid and carbohydrate content and the organic weight calculated as the sum of these components (Fig. 8B). There was a linear relationship between the scope for growth and fecundity. The weight of the eggs, however, did not reduce linearly with scope for growth. Over the range from $+5$ to -5 $\text{mg} \cdot \text{day}^{-1}$ in growth potential the weight of the eggs was similar at 73 ± 8.6 (SD) $\cdot 10^{-3}$ μg . Under more extreme stress the eggs were smaller.

The morphological processes that occurred in the gonads of mussels that were stressed during gametogenesis were complex. Resorption of morphologically ripe gametes occurred by autolysis (increased activity of lysosomal hydrolases within the oocytes) and by haemocytic infiltration of the gonad followed by phagocytosis. However, these processes occurred heterogeneously within the gonad tissue and, in addition, some gametocytes continued to develop within some follicles. At the end of 8 wk all individuals had some (albeit few) ripe gametes present in small follicles. The net effect was a reduction in fecundity under stress but a maintenance of the organic weight of the eggs, at least until the stress was considerable.

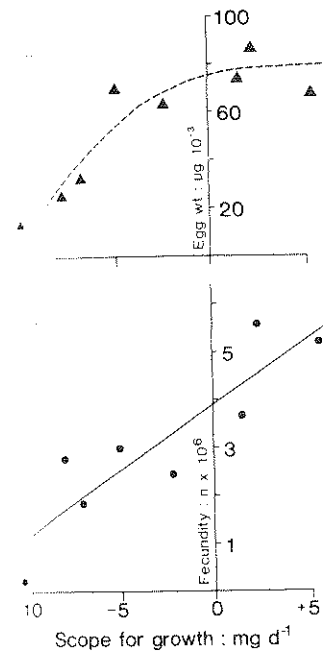


Fig. 8. A: numbers of eggs released by *Mytilus*; B: weight of eggs from the experiments shown in Fig. 8A.

Fig. 9. The residual reproductive value (RRV) versus age. Δ , Lynher population; \circ , Catterick population.

In order to relate these findings to the field, we studied a population of mussels in a polluted area, combined with poor rations during the winter and early spring (Bayne et al. 1978). As expected, fecundity was much lower. The fecundity per spawning was 2.8×10^5 $\text{eggs} \cdot \text{g}^{-1}$ in a nearby unpolluted area, compared with 14.2×10^5 $\text{eggs} \cdot \text{g}^{-1}$ in a nearby unpolluted area (Worrall and Iwama 1978). The data for fecundity and mortality were calculated as a fundamental component of the RRV to a value of 1.0 for the high age. The age-related RRV is much lower in the polluted area. In addition, the age of maximum RRV is further disadvantaged, since the age of maximum RRV in the breeding population ($< 10\%$ of the population) is much lower in the polluted area ($> 30\%$). Sub-lethal

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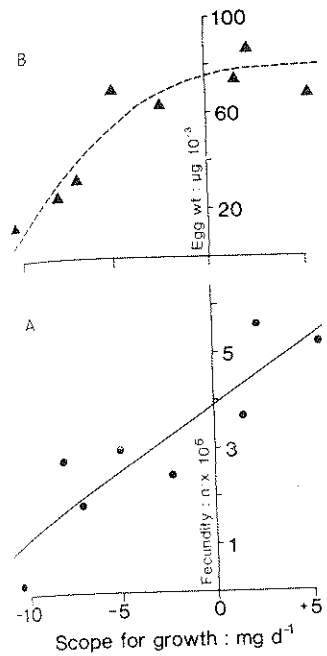


Fig. 8. A: numbers of eggs released by *Mytilus edulis* related to the scope for growth as mg · day⁻¹; B: the weight of eggs from the experiments shown in Fig. A.



Fig. 9. The residual reproductive value (RRV) of individual *Mytilus edulis* from two populations, related to age. Δ , Lynher population; \circ , Cattewater population (see Bayne and Worrall, 1980).

In order to relate these findings to conditions in the natural environment, we have studied a population of mussels in which unseasonally high winter water temperatures, combined with poor ration conditions, caused negative scope for growth in the winter and early spring (Bayne and Widdows, 1978; Bayne and Worrall, 1980). As expected, fecundity was much reduced in this population; estimated mean fecundity per spawning was 2.8×10^5 eggs per gram dry weight, compared with 14.2×10^5 eggs · g⁻¹ in a nearby population. We have also estimated mortality in both populations (Worrall and Bayne, unpubl. data). From age (= weight) related data for fecundity and mortality the residual reproductive value (RRV) can be calculated as a fundamental component of fitness (Fisher, 1930). The results, normalised to a value of 1.0 for the highest RRV in either population, are plotted in Fig. 9. Age-related RRV is much lower in the stressed population (the Cattewater). In addition, the age of maximum RRV increases in this population and this incurs further disadvantage, since these older individuals comprise a small proportion of the breeding population (<10%) compared with the age class of maximum RRV in the Lynher (>30%). Sub-lethal environmental stresses that reduce the individuals'

scope for growth clearly can have profound ecological consequences for the population.

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We have drawn liberally on the results, both published and unpublished of the Stress and Pollution group at I.M.E.R. and wish to thank J. Widdows, C.M. Worrall, P. Salkeld, D.M. Lowe, D. Dixon and S. Moore. Discussions with J. Widdows have helped to clarify our ideas. This work forms part of the Experimental Ecology Programme of the Institute for Marine Environmental Research, a component of the Natural Environment Research Council. It was funded in part by the Department of the Environment.

APPENDIX

Let y_{ij} denote the (log) rate from animal j in sample i , and x_{ij} its (log) weight, where $i = 1, 2$ and $j = 1, 2, \dots, n$. The three cases, and the null (H_0) and alternative (H_1) hypothesis examined are:

- $y_{ij} = \alpha_i + \text{'error'}$, $H_0: \alpha_2 = \alpha_1$, $H_1: \alpha_2 - \alpha_1 = \Delta\alpha$.
- $y_{ij} = \alpha_i + \beta x_{ij} + \text{'error'}$, $H_0: \alpha_2 = \alpha_1$, $H_1: \alpha_2 - \alpha_1 = \Delta\alpha$.
- $y_{ij} = \alpha_i + \beta_i x_{ij} + \text{'error'}$, $H_0: \alpha_2 + \beta_2 x_0 = \alpha_1 + \beta_1 x_0$, $H_1: (\alpha_2 + \beta_2 x_0) - (\alpha_1 + \beta_1 x_0) = \Delta\alpha$.

Here the 'error' is assumed normally distributed with constant variance σ^2 .

The test statistics T are functions of

$$\bar{y}_i = \sum_j y_{ij}/n, V(y_i) = \sum_j (y_{ij} - \bar{y}_i)^2/n, C(x_i, y_i) = \sum_j (x_{ij} - \bar{x}_i)(y_{ij} - \bar{y}_i)/n,$$

etc., namely:

- $T = (\bar{y}_2 - \bar{y}_1)/(2s^2/n)^{1/2}$, where $s^2 = [n/(2n-2)][\sum_i V(y_i)]$.
- $T = (\hat{\alpha}_2 - \hat{\alpha}_1)/(2k^2s^2/n)^{1/2}$ where $\hat{\alpha}_i = \bar{y}_i - \beta \bar{x}_i$ ($i = 1, 2$), $\beta = [\sum_i C(x_i, y_i)]/[\sum_i V(x_i)]$ and $s^2 = [n/(2n-3)][\sum_i V(y_i) - \beta^2 \sum_i V(x_i)]$, (for k see text).
- $T = [(\hat{\alpha}_2 + \hat{\beta}_2 \bar{x}_0) - (\hat{\alpha}_1 + \hat{\beta}_1 \bar{x}_0)]/(2k^2s^2/n)^{1/2}$ where $\hat{\alpha}_i = \bar{y}_i - \beta_i \bar{x}_i$, $\hat{\beta}_i = C(x_i, y_i)/V(x_i)$ ($i = 1, 2$) and $s^2 = [n/(2n-4)][\sum_i (V(y_i) - \hat{\beta}_i^2 V(x_i))]$, (for k see text).

Defining $t_v(P/2)$ to be the upper 100 $(P/2)\%$ point of the t distribution on v d.f., H_0 is rejected by a 2-tailed test of significance level P if

$$T > t_v(P/2) \text{ or } T < -t_v(P/2)$$

where (a) $v = 2n - 2$, (b) $v = 2n - 3$, (c) $v = 2n - 4$. t_v is the upper 100 $(P/2)\%$ point of the F distribution on $(1, v)$ d.f.

The power of such a test, against H_1 , is given by that

$$1 - \Phi(z(P/2) - c) + \Phi(-z(P/2) - c),$$

where $T_{v,\delta}$ has a non-central t distribution with v d.f. and non-centrality parameter $\delta = (\Delta\alpha)/(k\sigma)$. (See Scheffe (1959) for a non-central t distribution.) Note that c is determined through the 'scaled difference' $\Delta\alpha$.

The power for all experimental situations is determined by the non-central t distribution.

The non-central t distribution distribution tables are not available on computers. Computations are required as part of a computer program.

Values of P and n not covered by Fisher's (1930) tables are suggested:

Power $\approx 1 - \Phi(z(P/2) - c) + \Phi(-z(P/2) - c)$ where $z(P/2)$ is the upper 100 $(P/2)\%$ point of the $N(0, 1)$ distribution, $\Phi(x)$ is the $N(0, 1)$ distribution function.

where $z(P/2)$ is the upper 100 $(P/2)\%$ point of the $N(0, 1)$ distribution, $\Phi(x)$ is the $N(0, 1)$ distribution function.

where $z(P/2)$ is the upper 100 $(P/2)\%$ point of the $N(0, 1)$ distribution, $\Phi(x)$ is the $N(0, 1)$ distribution function.

$$c = \delta \left[1 - (z(P/2))^2 \left(\frac{1}{2v} + \frac{1}{8v^2} \right) \right]^{1/2}$$

The approximation relies only on the normal distribution and is adequate for $v \geq 8$ and very accurate for $v \geq 15$.

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ecological consequences for the

where (a) $v = 2n - 2$, (b) $v = 2n - 3$, (c) $v = 2n - 4$. Equivalently, T^2 can be referred to tables of the F distribution on $(1, v)$ d.f.

The power of such a test, against alternative H_1 , can be shown to be the probability that

$$T_{v,\delta} > t_v(P/2) \text{ or } T_{v,\delta} < -t_v(P/2),$$

where $T_{v,\delta}$ has a non-central t distribution on v d.f., with non-centrality parameter $\delta = (n/2)^{1/2}(\Delta\alpha)/(k\sigma)$. (See Scheffe (1959), Appendix IV, for a definition of the non-central t distribution.) Note that the power is a function of $\Delta\alpha$, k and σ^2 only through the 'scaled difference' $\Delta\alpha/(k\sigma)$; thus, for given P a single set of curves determines the power for all experimental conditions.

The non-central t distribution does not have a closed form and routines for its computation are not available on most computing systems. Thus if power calculations are required as part of a computer programme (or if curves are needed for values of P and n not covered by Figs. 6 and 7) the following simple approximation is suggested:

$$\text{Power} \approx 1 - \Phi(z(P/2) - c) + \Phi(-z(P/2) - c),$$

where $z(P/2)$ is the upper 100 $(P/2)\%$ point of the $N(0, 1)$ distribution (e.g. 1.96 if $P = 0.05$), $\Phi(x)$ is the $N(0, 1)$ distribution function, and

$$c = \delta \left[1 - (z(P/2))^2 \left(\frac{1}{2v} + \frac{1}{8v^2} \right) \right]^{1/2}.$$

The approximation relies only on widely available routines (or tables) for $\Phi(\cdot)$; it is adequate for $v \geq 8$ and very accurate for $v \geq 15$.

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with constant variance σ^2 .

$$(x_{ij} - \bar{x}_i)(y_{ij} - \bar{y}_i)/n,$$

$$[\sum_i V(y_i)].$$
$$1, 2), \hat{\beta} = [\sum_i C(x_i, y_i)] / [\sum_i V(x_i)]$$
$$)], \text{ (for } k \text{ see text).}$$
$$\hat{\alpha}_i = \bar{y}_i - \hat{\beta}_i x_i, \hat{\beta}_i = C(x_i, y_i) / V(x_i)$$
$$^2 V(x_i)], \text{ (for } k \text{ see text).}$$

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HYDROMINERAL BALANCE OF BROOK SALMO GAIRDNERI, AS AFFECTED BY PETROLEUM

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Rainbow trout, *Salmo gairdneri*, which was exposed to petroleum by a number of modes (aerosol, emulsion) for their effect on ion balance, osmolality and ATPase. Emulsion, using micron-size particulate emulsifier, was the most soluble component of the oils only, or emulsion exposure, evidenced by epithelial damage to gill lamellae. Other treatments had few effects. Ion concentrations (Na⁺, K⁺) were depressed in fresh water compared to control sea-water acclimated fish. Osmolality decreased in fresh water. Hydromineral imbalances were concluded from ion and ATPase imbalance and perhaps ATPase activity.

Key words: trout; salinity; petroleum;

INTRODUCTION

Much of the petroleum spill is in emulsion, or particulate form (oil droplets) and the action or the aerosol effect of emulsifiers is thought to counteract oil spills at sea. The action of emulsifiers on oil since emulsion formation is not clear. This study has been carried out to assess the effects of emulsifier concentrations, little definitively the effects of oil-in-water emulsions.

A specific and direct site of action for emulsifier is thought to be a number of physiologic

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